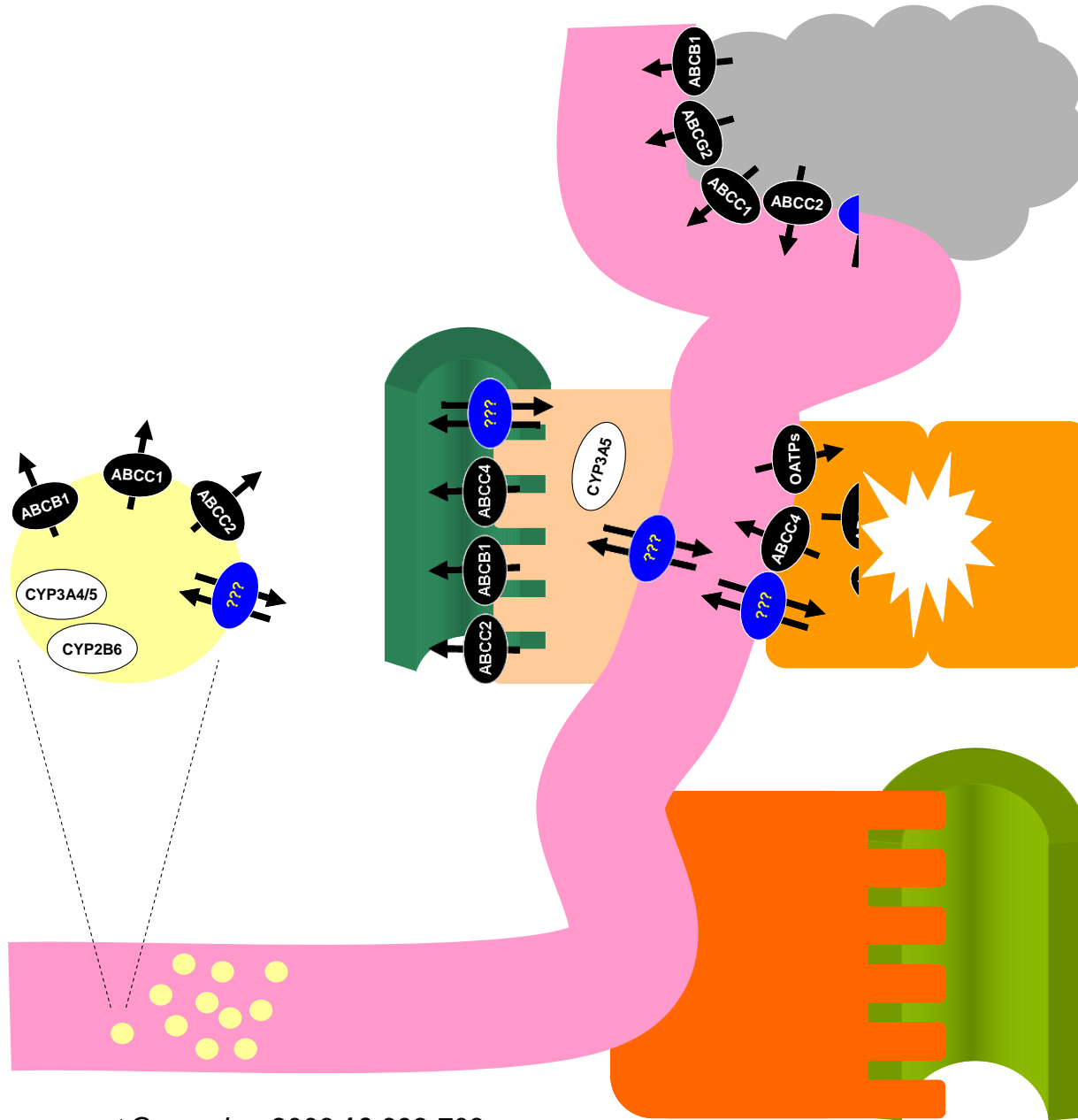


# The importance of studying drug transporters in HCV drug development



Andrew Owen, Ph.D.  
Professor of Pharmacology  
Department of Molecular and Clinical Pharmacology  
University of Liverpool, UK

# Coordinated drug transport and metabolism



# Mechanisms for hepatic residence and clearance of drugs

BLOOD

HEPATOCYTES

## Pharmacokinetics (PK)

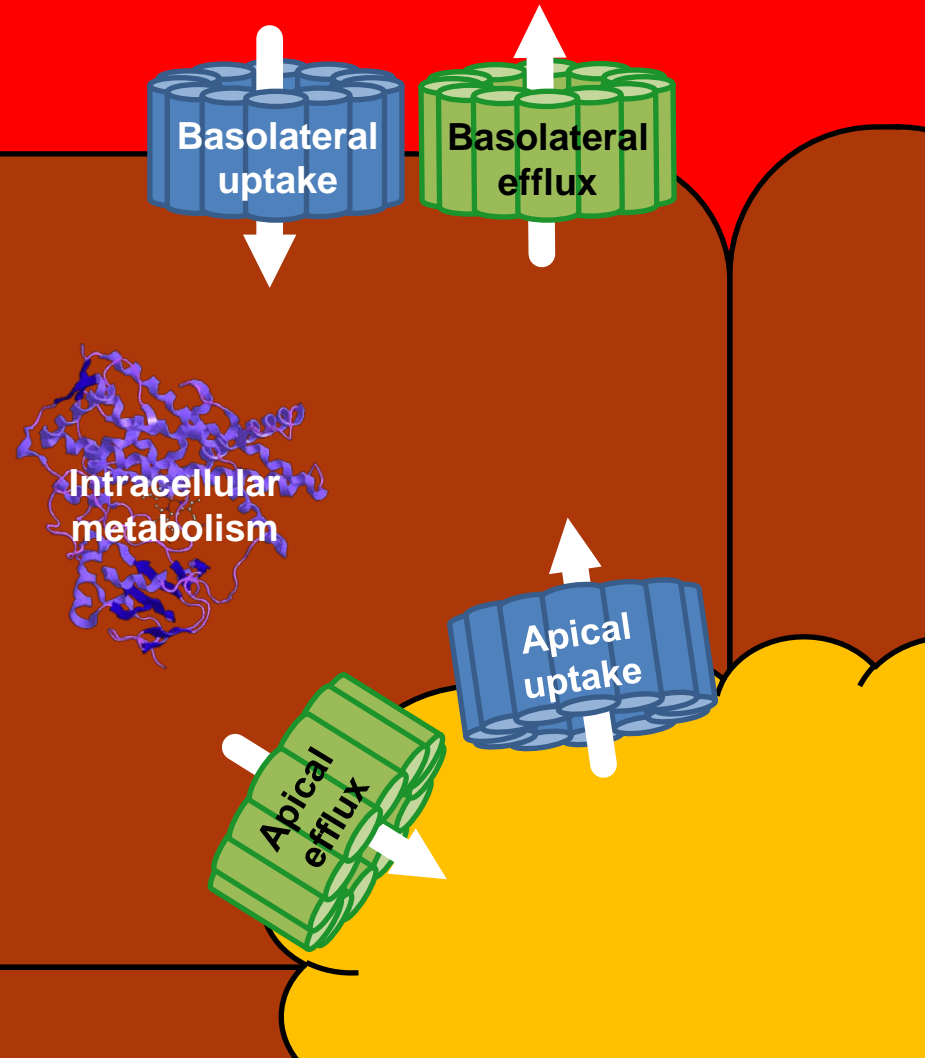
- Genetic/functional variability may translate to variability in plasma PK.
- Inhibition/induction by a “*perpetrator*” drug may affect PK of a “*victim*” drug.

## Hepatic intracellular concentrations (IC)

- Genetic/functional variability may influence IC.
- Inhibition/induction by a “*perpetrator*” drug may affect IC of a “*victim*” drug.

## Toxicity (hyperbilirubinemia)

- Genetic/functional variability may affect bilirubin uptake, conjugation or apical efflux..
- Inhibition/induction by a drug may affect bilirubin uptake, conjugation or apical efflux..



# International Transporter Consortium (ITC)

nature publishing group

REVIEW

## ITC Recommendations for Transporter Kinetic Parameter Estimation and Translational Modeling of Transport-Mediated PK and DDIs in Humans

MJ Zamek-Gliszczyński<sup>1</sup>, CA Lee<sup>2</sup>, A Poirier<sup>3</sup>, J Bentz<sup>4</sup>, X Chu<sup>5</sup>, H Ellens<sup>6</sup>, T Ishikawa<sup>7</sup>, M Jamei<sup>8</sup>, JC Kalvass<sup>9</sup>, S Nagar<sup>10</sup>, KS Pang<sup>11</sup>, K Korzekwa<sup>10</sup>, PW Swaan<sup>12</sup>, ME Taub<sup>13</sup>, P Zhao<sup>14</sup> and A Galetin<sup>15</sup>, on behalf of the International Transporter Consortium

This white paper provides a critical analysis of the importance of proper parameter calculation in various experimental settings, animal findings and application of static and dynamic models of human transporter-mediated pharmacokinetics to provide appropriate guidance for the use of transporter inhibitors in clinical science.

### IN VITRO TRANSPORTER KINETICS

The importance of active uptake and efflux in the intestine, kidney, and brain to drug pharmacokinetics has become increasingly more appreciated.<sup>1,2</sup> Understanding pharmacokinetics and drug–drug interaction (DDI) implications of transporter involvement in absorption, distribution, and excretion requires appropriate characterization of uptake and efflux *in vitro*. Two general types of *in vitro* systems are commonly used to study uptake and efflux transport kinetics: (i) experimental systems, including immortalized cell lines (e.g., CHO, HEK293, LLC-PK1, and MDCKII), oocytes, and vesicles, and (ii) primary and cultured cells (e.g., hepatocytes, primary derived cell lines (e.g., Caco-2, HepG2)). Expression of transporters can be directly used to estimate kinetic/inhibition parameters for the overexpressed transporter. By contrast, cell-based systems can be optimized to estimate kinetic parameters for uptake, metabolism, or efflux, as well as the interplay of multiple processes.

<sup>1</sup>Drug Disposition, Lilly Research Laboratories, Lilly Corporate Center, North Carolina, USA; <sup>2</sup>Non-Clinical Drug Safety, F. Hoffmann-La Roche, Pennsylvania, USA; <sup>3</sup>Department of Pharmacokinetics, Pharmacokinetics, GlaxoSmithKline Pharmaceuticals, King of Prussia, Pennsylvania, USA; <sup>4</sup>Medical Products Agency, Efficacy and Safety II, Uppsala, Sweden; <sup>5</sup>Office of Clinical Pharmacology, OTS, CDER, Food and Drug Administration, Silver Spring, USA; <sup>6</sup>Non-Clinical Safety, F. Hoffmann-La Roche Ltd., Switzerland; <sup>7</sup>Department of Pharmacokinetics, Pharmacodynamics and Drug Metabolism, Merck & Co., Rahway, USA; <sup>8</sup>Pharmacokinetics and Drug Metabolism, Pfizer Inc., Groton, USA; <sup>9</sup>Drug Disposition, Lilly Research Laboratories, Indianapolis, IN 46285, USA; <sup>10</sup>School of Pharmacy, University of Manchester, Manchester, UK; <sup>11</sup>Department of Pharmacokinetics, GlaxoSmithKline, Research Triangle Park, USA; <sup>12</sup>Drug Disposition, Lilly Research Laboratories, Indianapolis, IN 46285, USA; <sup>13</sup>Drug Disposition, Lilly Research Laboratories, Indianapolis, IN 46285, USA; <sup>14</sup>Drug Disposition, Lilly Research Laboratories, Indianapolis, IN 46285, USA; <sup>15</sup>School of Pharmacy, University of Manchester, Manchester, UK

Received 28 December 2012; accepted 21 February 2013; advance online publication 14 March 2013

CLINICAL PHARMACOLOGY & THERAPEUTICS

ACCEPTED ARTICLE PREVIEW

## Emerging Transporters of Clinical Importance: An Update from the International Transporter Consortium

Kathleen M. Hillgren<sup>1\*</sup>, Dietrich Keppler<sup>2\*</sup>, Arik Zur<sup>3</sup>, Kathleen M. Giacomini<sup>4</sup>, Bruno Stieger<sup>5</sup>, Carol E. Cass<sup>6</sup>, and Lei Zhang<sup>7\*</sup>

(on behalf of the International Transporter Consortium)

<sup>1</sup>Drug Disposition, Lilly Research Laboratories, Indianapolis, IN 46285, USA; Email: k.hillgren@lilly.com

ACCEPTED ARTICLE PREVIEW



INTERNATIONAL  
TRANSPORTER  
CONSORTIUM

### Transporter Studies in Drug Development: Experience to Date and Follow up on Decision Trees from the International Transporter Consortium

Donald Tweedie<sup>a</sup>, Joseph W. Polli<sup>b</sup>, Eva Gil Berglund<sup>c</sup>, Shiew Mei Huang<sup>d</sup>, Lei Zhang<sup>d</sup>, Agnès Poirier<sup>e</sup>, Xiaoyan Chu<sup>f</sup>, and Bo Feng<sup>g</sup>

<sup>a</sup>Drug Metabolism and Pharmacokinetics, Boehringer Ingelheim Pharmaceuticals Inc., Ridgefield, USA

<sup>b</sup>Drug Metabolism and Pharmacokinetics, GlaxoSmithKline, Research Triangle Park, USA

<sup>c</sup>Medical Products Agency, Efficacy and Safety II, Uppsala, Sweden

<sup>d</sup>Office of Clinical Pharmacology, OTS, CDER, Food and Drug Administration, Silver Spring, USA

<sup>e</sup>Non-Clinical Safety, F. Hoffmann-La Roche Ltd., Switzerland

<sup>f</sup>Department of Pharmacokinetics, Pharmacodynamics and Drug Metabolism, Merck & Co., Rahway, USA.

<sup>g</sup>Pharmacokinetics and Drug Metabolism, Pfizer Inc., Groton, USA.

### Introduction

The International Transporter Consortium organized a second Workshop in March 2012 to expand on the work of the first workshop and to address emerging transporters of clinical importance.

Dieter Seidelberg, Germany; Email: seidelberg@pharm.uni-wuerzburg.de

Michael J. Zamek-Gliszczyński, University of California, San Diego; Email: mjzamek@ucsf.edu; Tel: 415-476-1936

Kathleen M. Giacomini, University of California, San Diego; Email: kgiacomi@ucsf.edu; Tel: 415-476-1936  
Department of Pharmacology and Toxicology, 8091 Zurich, Switzerland; Tel: 41-76-634-3169

David W. Brown, National Cancer Institute, 11560 University Ave., Bethesda, MD, USA; Email: dwbrown@nih.gov  
David W. Brown, University of Alberta; Email: dwbrown@ualberta.ca; Tel: 780-436-4911  
David W. Brown, National Sciences, CDER, FDA, 10903 Newington, MD, USA; Email: dwbrown@fda.hhs.gov

© 2013 International Transporter Consortium. All rights reserved

# Transporters in regulatory documents

Transporter	Pseudonym(s)	ITC recommendation (Apr 2013)	FDA Guidance (Feb 2012)	EMA Guidelines (June 2012)
ABCB1	P-gp/MDR1	✓	✓	✓
ABCC2	MRP2	✓		
ABCG2	BCRP	✓	✓	✓
ABCB11	BSEP	✓		✓
SLCO1B1	OATP1B1	✓	✓	✓
SLCO1B3	OATP1B3	✓	✓	✓
SLC22A1	OCT1	✓		✓
SLC22A2	OCT2	✓	✓	✓
SLC22A6	OAT1	✓	✓	✓
SLC22A8	OAT3	✓	✓	✓
SLC47A1	MATE1	✓	✓	✓
SLC47A2	MATE2	✓	✓	✓

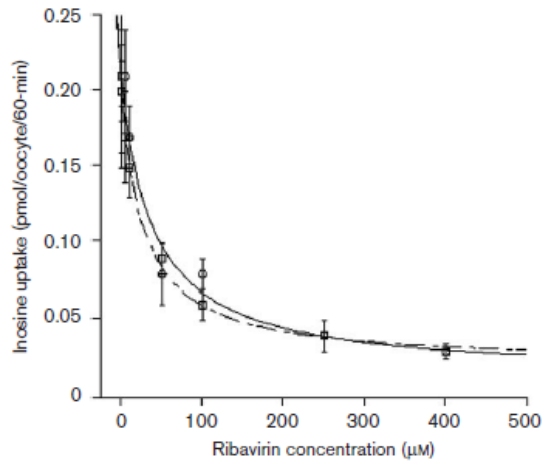
# SLC28 (CNT) and ribavirin

Original article 83

## Genetic analysis and functional characterization of polymorphisms in the human concentrative nucleoside transporter, CNT2

Ryan P. Owen<sup>a</sup>, Jennifer H. Gray<sup>a</sup>, Travis R. Taylor<sup>b</sup>, Elaine J. Carlson<sup>b</sup>, Conrad C. Huang<sup>b</sup>, Michiko Kawamoto<sup>b</sup>, Susan J. Johns<sup>b</sup>, Doug Stryke<sup>b</sup>, Thomas E. Ferrin<sup>b</sup> and Kathleen M. Giacomini<sup>a</sup>

Variant	Mean $\pm$ SD $K_m$ ( $\mu$ M)
CNT2-reference	35.5 $\pm$ 9.27
CNT2-P22L	40.8 $\pm$ 6.47
CNT2-S75R	31.2 $\pm$ 15.8
CNT2-S245T	26.7 $\pm$ 6.13
CNT2-F355S	49.9 $\pm$ 14.6



ORIGINAL ARTICLE

## Negative Predictive Value of *IL28B*, *SLC28A2*, and *CYP27B1* SNPs and Low RBV Plasma Exposure for Therapeutic Response to PEG/IFN-RBV Treatment

Antonio D'Avolio, BSc, MSc, SM,\* Alessia Ciancio, MD, PhD,† Marco Siccardi, BSc, MSc, PhD,‡ Antonina Smedile, MD,† Marco Simiele, BSc, MSc,\* Jessica Cusato, BSc, MSc,\* Lorena Baietto, BSc, MSc,\* Diego Aguilar Marucco, MD,\* Giuseppe Cariti, MD,\* Andrea Calcagno, MD,\* Daniel Gonzalez de Requena, BSc, MSc,\* Mauro Scudra, LT,\* Giulia Troshina, BSc, MSc,† Gian Paolo Caviglia, BSc, MSc,† Stefano Bonora, MD,\* Mario Rizzetto, MD,† and Giovanni Di Perri, MD, PhD\*

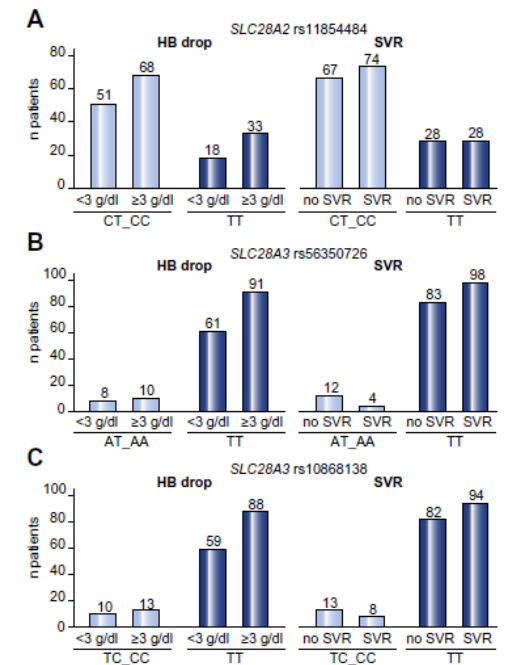
Factors	Multivariate	
	OR (95% CI)	P
Gender (male)	—	—
Weight (10 kg)	NS	NS
Age (10 yrs)	NS	NS
pegIFN-2a	—	—
RBV dose (mg/kg)	—	—
Dose reduction	—	—
RBV concentrations > 2.5 ( $\mu$ g/mL)	3.94 (1.43–10.91)	0.008
<i>IL28B</i> rs8099917TT	3.06 (1.14–8.1)	0.026
<i>CYP27B1</i> rs4646536TT	4.5 (1.54–13.1)	0.006
<i>CNT2</i> rs11854484TT	3.7 (1.3–10.83)	0.014

Research Article

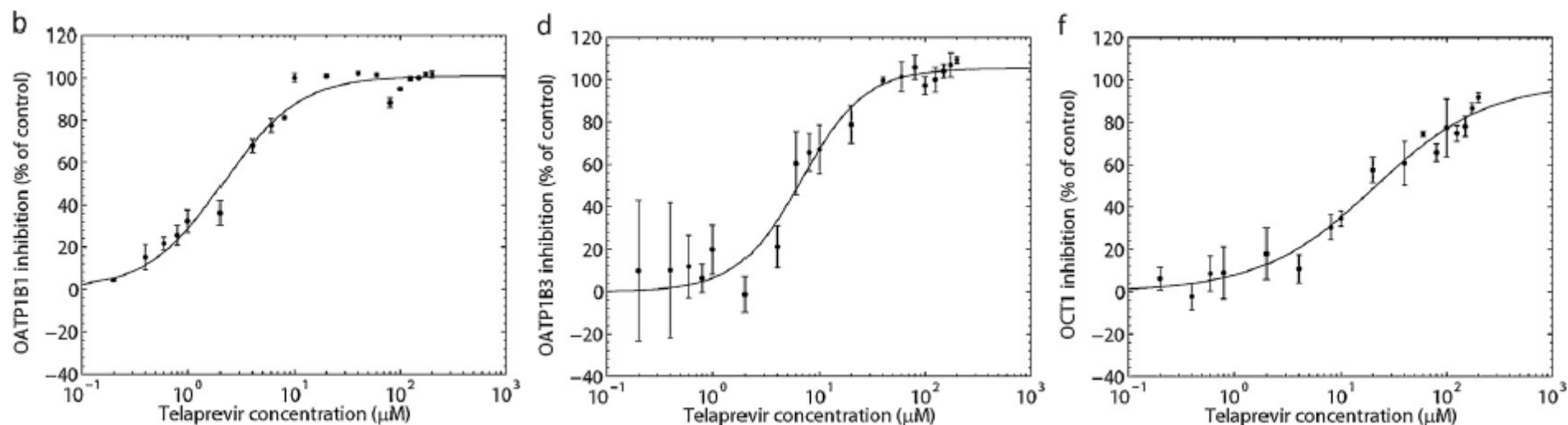
EASL JOURNAL OF HEPATOLOGY

Viral Hepatitis

## Impact of genetic *SLC28* transporter and *ITPA* variants on ribavirin serum level, hemoglobin drop and therapeutic response in patients with HCV infection



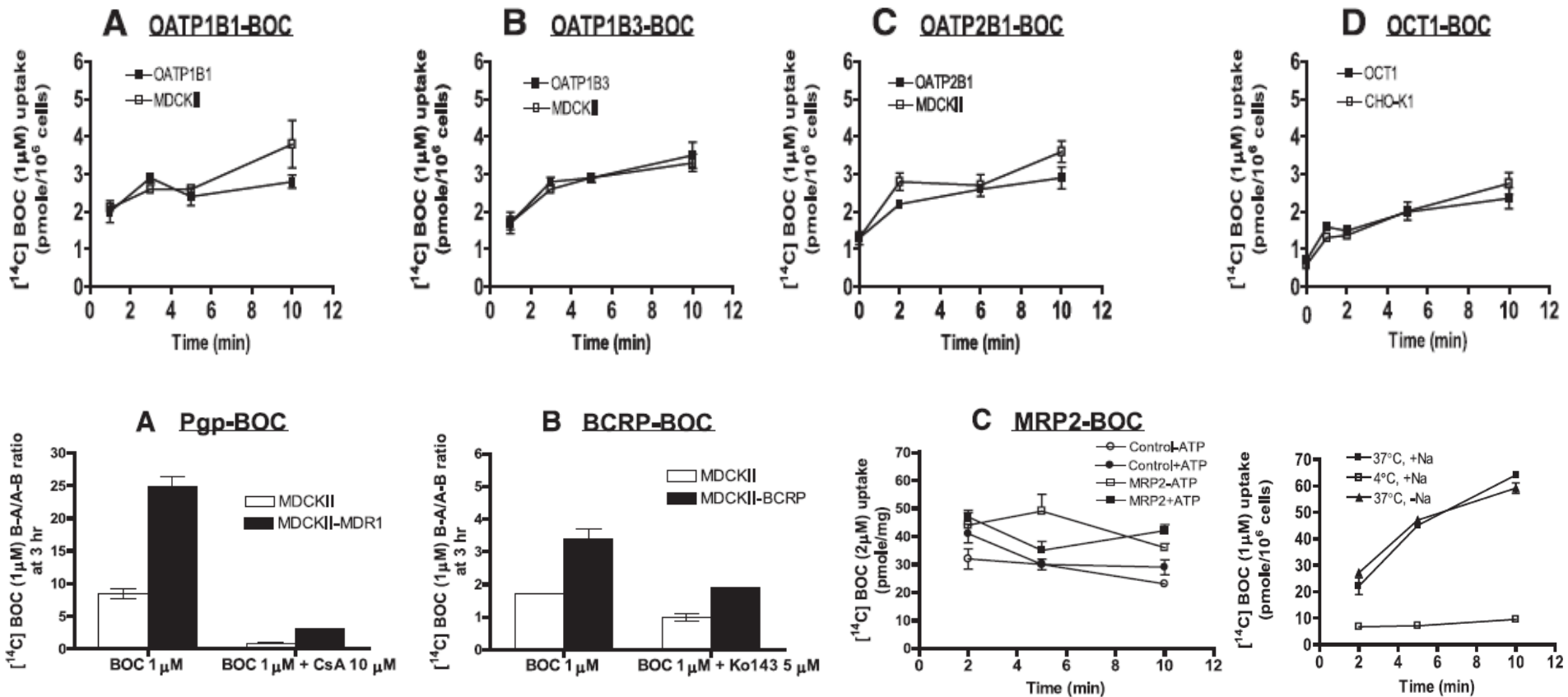
## Interaction of the antiviral drug telaprevir with renal and hepatic drug transporters



Localization	Transporter	IC <sub>50</sub>			% <sub>max</sub>		
		Value ( $\mu\text{M}$ )	SD ( $\mu\text{M}$ )	P	Value (%)	SD (%)	P
Kidney	OCT2	6.35	3.45	0.0856	62.29	6.91	<0.001
Kidney	MATE1	22.98	6.72	0.0033	111.76	9.95	<0.001
Liver	OATP1B1	2.15	0.13	<0.001	100.90	2.31	<0.001
Liver	OATP1B3	6.77	0.87	<0.001	105.39	3.88	<0.001
Liver	OCT1	20.67	7.74	0.0175	98.59	10.68	<0.001

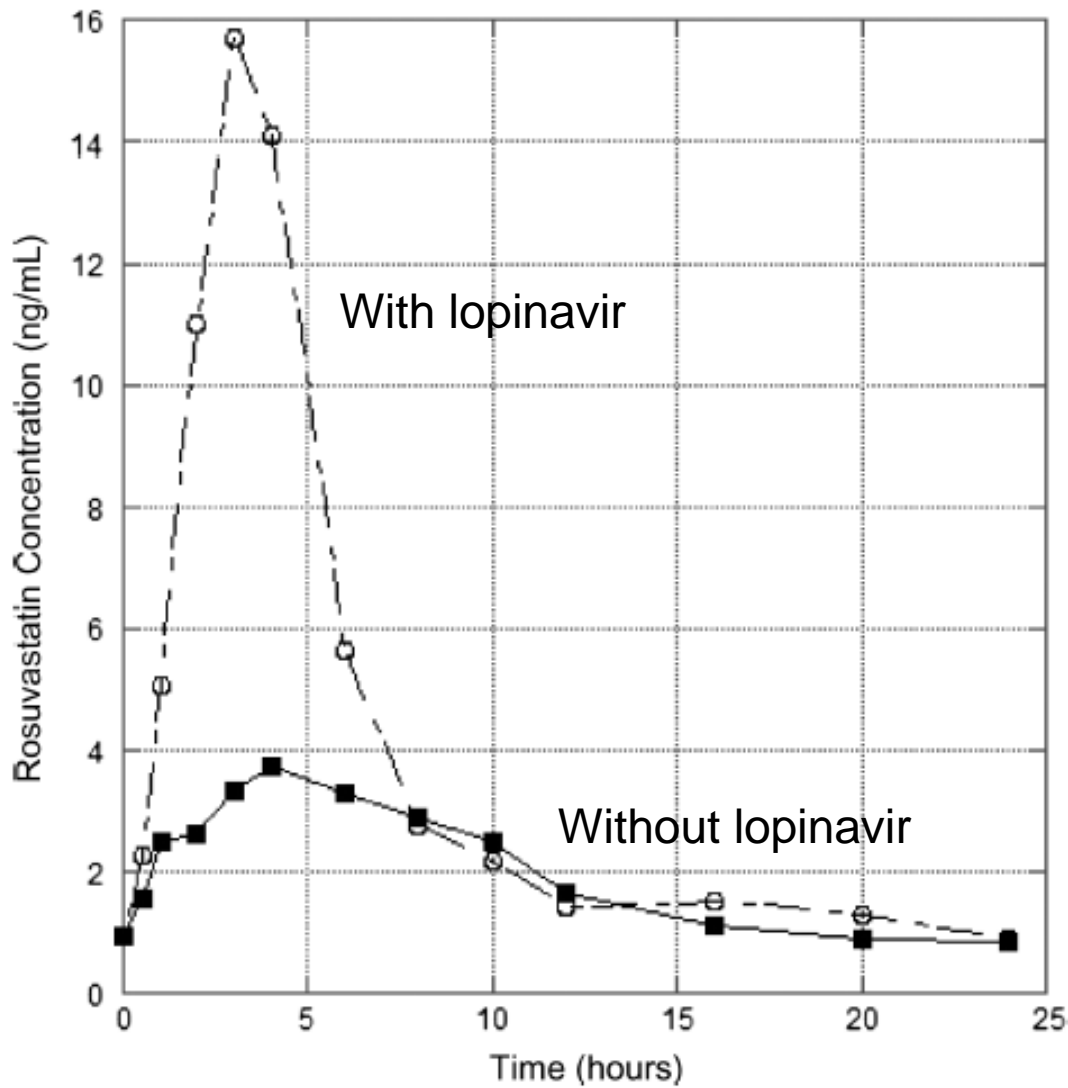
# In Vitro Assessment of Drug-Drug Interaction Potential of Boceprevir Associated with Drug Metabolizing Enzymes and Transporters<sup>S</sup>

Xiaoyan Chu, Xiaoxin Cai, Donghui Cui, Cuyue Tang, Anima Ghosal, Grace Chan, Mitchell D Green, Yuhsin Kuo, Yuexia Liang, Cheri M Maciolek, Jairam Palamanda, Raymond Evers, and Thomayant Prueksaritanont





# In Vitro Assessment of Drug-Drug Interaction Potential of Rosuvastatin Associated with Drug Metabolizing Enzymes



Kiser et al. *J Acquir Immune Defic Syndr*. 2008 Apr 15;47(5):570-8.

Xiao  
Mits

Chan,  
amanda,

Perpetrator

References

BOC (800 mg, thr  
CsA (100 mg)  
Rifampin (600 mg)  
Lopinavir (400 mg)  
Telaprevir (750 mg)  
Amprenavir (600 mg)  
Ritonavir (100 mg)

skotte EG et al., 2011  
ara and Sugiyama, 2006  
et al., 2009  
er et al., 2008  
et al., 2011  
gren et al., 2012  
ti et al., 2008

# HCV, liver disease and transporters



Pharmacological Reports  
2012, 64, 927-939  
ISSN 1734-1140

Copyright © 2012  
by Institute of Pharmacology  
Polish Academy of Sciences

Drug Metab. Pharmacokinet. 25 (2): 190-199 (2010).

## Regular Article

### Hepatitis C Virus-related Cirrhosis is a Major Determinant of the Expression Levels of Hepatic Drug Transporters

	mRNA levels (amol/ $\mu$ g total RNA)				Ratio [(+), (+)/(−), (−)]			
	HCV (−), LC (−)	HCV (+), LC (−)	HCV (−), LC (+)	HCV (+), LC (+)				
OCT1	16.63 ± 8.91	13.64 ± 9.23	10.70 ± 4.60	8.87 ± 5.56***	0.53			
OCTN2	0.70 ± 0.32	0.83 ± 0.47	0.55 ± 0.24	0.75 ± 0.39	1.07			
OAT2	11.86 ± 7.36	13.37 ± 8.62	10.06 ± 4.03	10.27 ± 5.66	0.87			
OAT7	0.62 ± 0.33	0.56 ± 0.30	0.47 ± 0.19	0.47 ± 0.20	0.76			
OATP1B1	10.55 ± 6.71	8.04 ± 4.78	6.72 ± 2.95	5.91 ± 2.89***	0.56			
OATP1B3	3.83 ± 2.27	2.91 ± 1.95	2.80 ± 1.54	1.98 ± 1.15**	0.52			
OATP2B1	31.20 ± 13.72	32.50 ± 20.18	24.13 ± 10.17	24.36 ± 10.91	0.78			
MATE1	3.14 ± 2.04	2.13 ± 1.27	2.04 ± 0.93	1.34 ± 0.68***	0.43			
PEPT1	1.01 ± 0.55	1.24 ± 0.66	0.81 ± 0.39	0.94 ± 0.32	0.93			
MDR1	1.15 ± 0.52	1.56 ± 0.87	1.07 ± 0.35	1.40 ± 0.43*	1.22			
MRP1	0.25 ± 0.14	0.40 ± 0.26	0.27 ± 0.17	0.51 ± 0.53*	2.04			
MRP2	2.96 ± 1.77	2.44 ± 1.66	1.45 ± 0.68	1.45 ± 0.54***	0.49			
MRP3	2.58 ± 1.31	2.64 ± 1.34	1.68 ± 0.55	2.18 ± 1.09	0.85			
MRP4	0.09 ± 0.05	0.16 ± 0.14	0.11 ± 0.05	0.18 ± 0.13***	2.06			
MRP5	0.07 ± 0.04	0.10 ± 0.06	0.06 ± 0.02	0.11 ± 0.08	1.49			
MRP6	3.41 ± 2.16	3.13 ± 1.76	2.50 ± 1.28	2.33 ± 1.15	0.68			
BCRP	0.57 ± 0.33	0.45 ± 0.25	0.33 ± 0.17	0.31 ± 0.22***	0.53			
			MRP5	0.07 ± 0.04	0.10 ± 0.06	0.06 ± 0.02	0.11 ± 0.08	1.49
			MRP6	3.41 ± 2.16	3.13 ± 1.76	2.50 ± 1.28	2.33 ± 1.15	0.68
			BCRP	0.57 ± 0.33	0.45 ± 0.25	0.33 ± 0.17	0.31 ± 0.22***	0.53

E  
A  
N  
B  
A  
C

3  
1

GSTA1  
SULT1A1  
SULT2A1  
UGT1A1  
UGT1A4  
UGT1A6  
ABCB1  
ABCC2  
ABCC3  
ABCC4  
ABCG2

MRP5 0.07 ± 0.04 0.10 ± 0.06 0.06 ± 0.02 0.11 ± 0.08 1.49  
MRP6 3.41 ± 2.16 3.13 ± 1.76 2.50 ± 1.28 2.33 ± 1.15 0.68  
BCRP 0.57 ± 0.33 0.45 ± 0.25 0.33 ± 0.17 0.31 ± 0.22\*\*\* 0.53

# Investigational (Phase 2+) HCV DAAs

Protease Inhibitors	
<b>BI 201335 (Faldaprevir)</b>	CYP3A substrate Moderate inhibitor of CYP3A
<b>TMC435 (Simeprevir)</b>	CYP3A substrate Weak inhibitor of CYP3A and P-gp
<b>ABT-450</b>	CYP3A substrate Boosted by ritonavir
<b>BMS-650032 (Asunaprevir)</b>	Weak inhibitor of CYP2D6 and P-gp Weak inducer of CYP3A4
<b>RG7227 (danoprevir)</b>	CYP3A substrate boosted by ritonavir

# Investigational (Phase 2+) HCV DAAs

Non-nucleoside polymerase inhibitors	
BI 207127	No data
Filibuvir (PF-868554)	CYP3A substrate Weak inducer and inhibitor
GS-9190 (Tegobuvir)	Metabolism (limited) by CYP1A2 No CYP inhibition and induction
ART-333	Metabolised by CYP2C8, 3A4, 2D6

# Investigational (Phase 2+) HCV DAAs

## NS5A Inhibitors

**BMS-790052 (daclatasvir)**

CYP3A substrate; P-gp substrate

P-gp inhibitor

## Nucleoside polymerase inhibitors

**GS-7977 (sofosbuvir)**

Not a CYP3A substrate

Renally excreted

# Summary and conclusions

- Transporters play a key role for inter-patient variability in pharmacokinetics, distribution, drug interactions and safety
- Robust interpretation of transporter data relies on knowledge of tissue and cellular transporter localisation which may be influenced by disease
- Over 400 transporters are coded within the human genome and relatively few have been studied for xenobiotic transport
- There is a paucity of published literature on the interaction of HCV drugs with transporters

# Acknowledgments

David Back  
Marco Siccardi  
Steve Rannard  
Saye Khoo  
Alessandro Schipani  
Paul Curley  
Deirdre Egan  
Phil Martin  
Neill Liptrott  
Lee Tatham  
Laura Dickinson  
Laura Else  
Sara Gibbons  
Helen Reynolds  
Kay Seden  
Vicky Watson  
Adeniyi Olagunju  
Beth Williamson  
James Hobson

South Africa  
Helen McIlleron  
Alexander Pym

Torino  
Giovanni Di Perri  
Stefano Bonora  
Andrea Calcagno  
Antonio D'Avolio

Barcelona  
Pepe Molto  
Marta Valle  
Bonaventura Clotet

Cologne  
Christoph Wyen  
Gerd Fätkenheuer

London  
Marta Boffito  
Mike Youle  
Margaret Johnson  
Akil Jackson

## Funding

